

Detection of low-dose irradiation of dry fruits by termoluminescence

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Abstract. The aim of the study was to investigate whether it is possible to detect low-dose irradiation in dried fruits using the thermoluminescence (TL) method. Selected dried fruits: strawberries, cherries, black currants, two types of raisins, mulberries, apricots, figs, dates and plums were irradiated with doses of 0.1 kGy, 0.3 kGy and 0.5 kGy.

Keywords: Dried fruits • Low doses of ionizing radiation • Thermoluminescence method

Introduction

This paper describes the possibility of using the thermoluminescence (TL) method for the detection of the irradiation of dried fruits using low doses of radiation. Dried fruits are used on an industrial scale as an active ingredient in phytopharmaceuticals, mainly in food supplements and in teas and herbal infusions. Market research on dried fruit shows that the number of consumers of these products is constantly growing [1, 2], and they should therefore meet the requirements to be considered as healthy and safe food.

Dried fruits, along with spices and herbs, belong to a group of products that are relatively frequently exposed to ionizing radiation for preservation purposes and to eliminate the incidental presence of pathogenic microorganisms. The recommended dose for preservation is approximately 10 kGy [3–12]. However, doses <10 kGy are often used for sanitary protection. According to Cruz-Zaragoza *et al.* [13], doses <0.4 kGy are used in many countries.

Despite the many concerns about the adverse effects prevailing, it can be pointed out that irradiation does not cause radioactivity in food; does not change the composition of food; does not reduce the nutritional value; and does not change the taste, texture, or appearance of food [14–21].

The European Committee for Standardization has published the standard EN 1788:2001 [22] for the detection of radiation by TL in foods from which silicate minerals can be isolated. TL is currently the most widely used method for detecting irradiation of seafood, spices, and herbs and their mixtures

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0029-5922 © 2024 The Author(s). Published by the Institute of Nuclear Chemistry and Technology. This is an open access article under the CC BY-NC-ND 4.0 licence (http://creativecommons.org/licences/by-nc-nd/4.0/). [23–49]. This method is also used for the detection of irradiation of fresh and dried fruits and vegetables [26–28, 31–35, 50–66].

Electron paramagnetic resonance (EPR) is also used to detect irradiation of dried fruits [67–80]. However, the EPR signals of irradiated fruits are complex and relatively broad, which can have a negative effect on the sensitivity of the method and therefore the difficulty of detecting irradiation of dried fruits with low doses of radiation. In the present work, 10 dried fruits were selected to study whether the irradiation treatment is detected by TL.

Experimental

In the present work, 10 types of dried fruits (strawberries, cherries, black currants, two kinds of raisins, mulberries, apricots, figs, dates, and plums) have been selected to study the detection of the irradiation treatment by TL. The products were purchased from Warsaw, Poland.

All chemicals of the analytical grade were purchased from commercial sources: sodium polytungstate (3 Na₂WO₄·9 WO₃·H₂O) was purchased from Sometu, Germany; hydrochloric acid (HCl), ammonium hydroxide (NH₃(aq), and acetone (C₃H₆O) were purchased from Avantor Performance Materials, Poland S.A.; and silicone spray was purchased from Henax Sp. z o.o., Poland. Nitrogen (N₂) for flushing the TL heating chamber was purchased from Air Products and Chemicals, Inc., with a purity of 99.98%. The water was at grade 3.

Gamma irradiation of dried fruits was performed in the Gamma Chamber GC5000 cobalt source (BRIT, India – installed at the Institute of Nuclear Chemistry and Technology) at doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy at a dose rate of 1.6 kGy/h. The absorbed dose was controlled with an alanine dosimeter according to the ISO/ASTM 51607:2013 (E) Standard Practice for use of an alanine-EPR dosimetry system [81].

Procedure of isolation of silicate minerals

The description of the testing methodology is based on the EN 1788:2001 standard [22]. The isolated silicate minerals are required to be free of organic material. The presence of organic matter could induce spurious (non-radiation induced) luminescence, or in extreme cases, could obscure TL. Samples with organic residues are found blackened by the TL measurement process [22].

Preconcentration step of minerals from the dried fruits

Preconcentration of minerals by wet sieving was performed for dried fruits. Samples weighing 200–300 g were placed in 1000 ml glass beakers, and water was added to cover the dried fruits.

The dried fruits in the beakers were treated with ultrasound for about 5 min (to loosen the adhering minerals).

Then, the dried fruits were sieved in portions through a 250- μ m nylon mesh into large beakers, rinsing the minerals through with water each time, using a strong jet of water from a wash bottle. The constituents retained by the sieve cloth were discarded. Solutions with the minerals were allowed to settle for 10 min.

Most of the water from the large beaker was decanted together with the organic minerals, leaving only the silicate minerals in a few millilitres of water. The mineral fraction was transferred to a centrifuge tube using a Pasteur pipette. The centrifuge tubes were centrifuged for 1 min at a centrifugal acceleration of 1000 g. The water was removed by suction, leaving the mineral fraction behind.

Density separation step to free the minerals from the organic material

To the mineral fraction in the centrifuge tubes was added 5 ml of sodium polytungstate solution with a density of 2 g/ml. Then, the tubes were shaken vigorously and agitated in an ultrasonic bath for 10 min.

The mineral fraction in the centrifuge tubes was centrifuged for 2 min at a centrifugal acceleration of 1000 g. The silicate minerals were sedimented, whereas the organic components were found floating.

Then, carefully the sodium polytungstate solution was overlayed with water to facilitate removal of the organic material. The upper water layer and the organic materials were extracted by vacuum suction, leaving the minerals behind in the lower polytungstate layer. If not all organic material is removed, overlayed the sodium polytungstate solution with water again and repeated the extraction.

The minerals were washed twice to remove the sodium polytungstate residues by filling the centrifuge tube with water and allowing the minerals to settle. The water was later removed.

To dissolve carbonates adhering to the silicate minerals, added 1 ml of hydrochloric acid of 3 mol/l and agitated and left for 10 min.

The acid was neutralized using an ammonium hydroxide solution of 1 mol/l, the centrifuge tube filled up with water, and allowed the minerals to settle. The supernatant fluid was removed, and the mineral residue was washed twice with water.

To displace the residual water, added about 3 ml of acetone and agitated. If the acetone became turbid, it was removed and a fresh portion of acetone was added.

Fixing the minerals on discs for TL measurement

The isolated silicate minerals in acetone were transferred to a disc using a Pasteur pipette. Stainless steel discs had a diameter of 9 mm and a thickness of 0.25 mm. The deposit of minerals was fixed on the disc by using silicone spray.

The discs with the silicate minerals were stored overnight at $50 \pm 5^{\circ}$ C in a laboratory oven. The acetone dried off leaving a deposit of silicate minerals adhering to the discs.

TL measurements

TL measurements of isolated silicate minerals were carried out with a TL reader, type TL/OSL, model TL-DA-20 (Risø National Laboratory, Denmark) under the measurement conditions recommended

by EN 1788:2001 standard [22]: Initial temperature: 70°C; heating rate: 6°C/s; preheat time: 30 s; final temperature: 450°C.

The sample was preheated to a temperature of about -70° Ĉ using the heating rate -6° C/s. Then, this temperature was held for 30 s. A temperature was run up to a temperature of -450° C.

Recording of the TL glow curve

The results of the TL measurements are recorded in the form of TL curves expressing the relationship between TL intensity and heating temperature registered in the range of 70-450°C. Finding of a maximum (peak) in the TL curve in the temperature range of 150-250°C is an evidence of sample Table 1. Results of TL measurements on dried fruits, nonirradiated and irradiated at doses 0.1 kGy, 0.3 kGy, and 0.5 kGy

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1	2	4	5	6	7	8
Product name	Dose (kGv)	TL intensity Glow 1 (150–250°C)	TL intensity Glow 2 (150–250°C)	Glow 1/Glow 2	TL max. Glow 1 (°C)	TL max. Glow 2 (°C)

Product name	Dose (kGy)	(150–250°C) (counts/s)*	Glow 2 (150–250°C) (counts/s)*	Glow 1/Glow 2	Glow 1 (°C)	Glow 2 (°C)
Dried strawberries	0	198 506	70 836 640	0.003	274	184
	0.1	4 523 523	65 911 546	0.069	189	185
	0.3	17 255 429	60 080 576	0.287	187	184
	0.5	12 389 562	25 769 557	0.481	184	185
Dried cherries	0	2 767	4 512 059	0.001	>300	189
	0.1	366 090	7 151 444	0.051	184	185
	0.3	1 038 791	4 650 785	0.223	182	178
	0.5	6 218 121	17 305 999	0.359	187	184
Dried blackcurrants	0 0.1 0.3 0.5	34 528 2 939 176 38 302 937 38 302 937	53 734 996 41 324 465 150 752 558 86 004 147	$0.001 \\ 0.071 \\ 0.254 \\ 0.445$	>300 189 176 182	190 185 178 189
Sultana raisins	0	10 854	7 716 995	0.001	>300	193
	0.1	626 748	13 023 415	0.048	202	195
	0.3	2 097 384	13 172 576	0.159	204	193
	0.5	3 007 242	8 762 420	0.343	202	189
Dried mulberry	0	1 950	1 780 820	0.001	>300	185
	0.1	490 265	6 362 997	0.077	215	199
	0.3	522 540	2 393 365	0.218	199	197
	0.5	662 538	247 763	0.267	208	197
Iranian raisins	0	30 023	33 247 193	0.001	295	189
	0.1	3 422 594	51 448 440	0.066	208	195
	0.3	7 404 863	41 464 001	0.178	202	187
	0.5	10 998 191	38 722 815	0.284	195	184
Dried apricots	0	3 738	4 129 258	0.001	>300	189
	0.1	114 759	23 246 668	0.049	200	197
	0.3	731 068	4 452 959	0.164	206	193
	0.5	810 285	2 946 271	0.275	200	193
Dried figs	0	12 859	4 621 315	0.003	>300	187
	0.1	236 994	6 997 634	0.034	200	191
	0.3	781 721	4 162 217	0.188	200	195
	0.5	1 012 569	3 444 112	0.294	189	184
Dried dates	0	2 588	7 102 126	0.001	>300	180
	0.1	945 821	19 370 203	0.049	200	189
	0.3	4 105 175	9 817 495	0.418	199	185
	0.5	1 106 343	2 940 401	0.376	193	185
Dried plums	0	12 627	7 551 362	0.002	300	197
	0.1	1 493 161	21 356 765	0.070	191	176
	0.3	2 617 091	13 326 800	0.196	202	182
	0.5	6 120 437	18 600 863	0.329	191	187

*Counts/s – number of light pulses registered in the photomultiplier of the TL reader.

After irradiation of the discs, they were stored overnight at $50 + 5^{\circ}$ C in a laboratory oven before recording Glow 2. Glow 2 was measured under the same conditions as Glow 1.

Results and discussion

The conducted research aimed to determine whether the TL method is effective in detecting irradiation of dried fruits with lower doses of ionizing radiation than the previously used doses in the range of 1–10 kGy.

The results obtained from measuring the TL of the dried fruits studied are presented in Table 1 (before irradiation – dose of 0 kGy and after irradiation – dose of 0.1 kGy, 0.3 kGy, and 0.5 kGy).

According to the EN 1788:2001 standard, in the TL method, a reliable proof of food irradiation is the analysis of TL glow curves and the following two conditions:

- Condition 1: The TL glow ratio (Glow1/Glow2) measured in the temperature range of 150–250°C must be >0.1 (see Table 1, column 6).
- Condition 2: The TL Glow 1 curve must show a clear maximum in the temperature range of 150–250°C (see Table 1, column 7).

From the 10 samples of remaining dried fruits (strawberries, cherries, blackcurrants, two kinds of raisins, mulberries, apricots, figs, dates, and plums), it was possible to isolate appropriate amounts of silicate minerals using the density method with sodium polytungstate solution. Samples of the 10 listed fruits irradiated with doses of 0.3 kGy and 0.5 kGy were unequivocally identified as irradiated. On the TL glow curves of the samples irradiated with these doses, characteristic peaks of glow could be distinguished in the temperature range from 176°C to 206°C (see Table 1, column 7), ideally within the temperature range of 150–250°C, in which according to the norm, only the glow of the irradiated samples is observed. Also, the TL Glow1/Glow2 ratios of the samples irradiated with these doses were higher than the critical value of 0.1. In the case of samples irradiated with a dose of 0.3 kGy, the ration were within the range of 0.159–0.418, while for samples irradiated with a dose of 0.5 kGy, the ratios were within the range of 0.267–0.481 (see Table 1, column 6).

Not so unambiguous results were obtained when examining samples irradiated with the lowest dose 0.1 kGy. The TL Glow1/Glow2 ratios for these samples deviated from 0.1 and were slightly lower, ranging from 0.034 to 0.077 (see Table 1, column 6).

On the other hand, in the TL glow curves of these samples, it was possible to distinguish peaks in the range of 150–250°C, recorded at temperatures from 184°C to 208°C (see Table 1, column 7 and Figs. 1–10), i.e., within in the range of 150–250°C, typical for irradiated samples.

Moreover, the EN 1788:2001 standard states that a sample has been irradiated if the ratio of glow 1 to Glow 2 is <0.1 and a clearly defined Glow 1 is observed in the range of 150–250°C. This is sufficiently reliable evidence of irradiation of the sample [22].

The TL Glow 1 records obtained with dried fruits (strawberries, cherries, blackcurrants, Sultana raisins, mulberries, Iranian raisins, apricots, figs, dates, and plums) irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy are shown in Figs. 1–10, respectively.



Fig. 1. TL Glow 1 records obtained with dried strawberries irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy of gamma radiation.



Fig. 2. TL Glow 1 records obtained with dried cherries irradiated with doses of 0.1 kGy, 0.3 kGy, 0.5 kGy of gamma radiation.



Fig. 3. TL Glow 1 records obtained with dried blackcurrants irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy of gamma radiation.



Fig. 4. TL Glow 1 records obtained with Sultana raisins irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy of gamma radiation.



Fig. 5. TL Glow 1 records obtained with dried mulberries irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy of gamma radiation.



Fig. 6. TL Glow 1 records obtained with Iranian raisins irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy of gamma radiation.



Fig. 7. TL Glow 1 records obtained with dried apricots irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy of gamma radiation.



Fig. 8. TL Glow 1 records obtained with dried figs irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy of gamma radiation.



Fig. 9. TL Glow 1 records obtained with dried dates irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy of gamma radiation.



Fig. 10. TL Glow 1 records obtained with dried plums irradiated with doses of 0.1 kGy, 0.3 kGy, and 0.5 kGy of gamma radiation.

Summary

The tests of dried fruits described in this paper carried out using the TL method showed the usefulness of this method to identify the irradiation of a selected group of dried fruits irradiated with doses lower than those recommended in normative documents.

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References

- Vantage Market Research & Consultancy Services. (2022). Dry Fruits Market – Global Industry Assessment & Forecast. Retrieved June 15, 2023, from https:// www.vantagemarketresearch.com/industry-report/ dry-fruits-market-1559.
- Research and Markets. (2023, February). Dried Fruits Global Market Report 2023. Retrieved June 15, 2023, from https://www.researchandmarkets.com/report/ dried-fruit?gclid=EAIaIQobChMIx6bwvMrE_wIVBwd7Ch3YfAjVEAAYASAAEgJLsfD_BwE.
- Kolek, Z. (2011). The use of ionizing radiation to preserve food. *Zeszyty Naukowe Uniwersytetu Ekonomicznego w Krakowie*, 874, 45–57. https://r.uek. krakow.pl/jspui/handle/123456789/232.
- Migdał, W., Gryczka, U., Bertrandt, J., Nowicki, T., & Pytlak, R. (2014). Radiation methods in decision support system for food safety. *Nukleonika*, 59(4), 161–168. DOI: 10.2478/nuka-2014-0022.
- Guzik, G. P., & Michalik, J. (2021). European intercomparison studies as a tool for perfecting irradiated food detection methods. *Nukleonika*, 66(3), 91–97. DOI: 10.2478/nuka-2021-0013.
- US Food and Drug Administration. (2018, February). Food irradiation: What you need to know. Retrieved June 15, 2023, from https://fda.gov/food/buy-storeserve-safe-food/food-irradiation-what-you-need-know.
- FAO/WHO. (1984). Codex general standard for irradiated foods and recommended international code of practice for the operation of radiation facilities used for the treatment of foods. In *Codex Alimentarius* (Vol. XV). Rome: Codex Alimentarius Commission.
- 8. World Health Organization. (1988). *Food irradiation: A technique for preserving and improving the safety of food*. Geneva, Switzerland: WHO.
- World Health Organization. (1999). High dose irradiation: Wholesomeness of food irradiated with doses above 10 kGy. Geneva, Switzerland: WHO. (WHO Technical Report Series 890).
- FAO/WHO. (2005). Fruit and vegetable for health. Report of a Joint FAO/WHO Workshop, Kobe, Japan, 2004. Kobe, Japan: World Health Organization and Food and Agriculture Organization of the United Nations.
- 11. European Commission. (1999). Directive 1999/2/EC of the European Parliament and of the Council on the approximation of the laws of the Member States concerning foods and food ingredients treated with ionizing radiation. EU.
- 12. European Commission. (1999). Directive 1999/3/EC of the European Parliament and of the Council on the establishment of a Community list of foods and food ingredients treated with ionizing radiation. EU.
- Cruz-Zaragoza, E., Marcazzó, J., & Chernov, V. (2012). Photo- and thermally stimulated luminescence of polyminerals extracted from herbs and spices. *Radiat. Phys. Chem.*, 81(8), 1227–1231. DOI: 10.1016/j.radphyschem.2012.01.024.
- European Food Safety Authority. (2011). Scientific opinion on the chemical safety of irradiation of food. *EFSA Journal*, 9(10), 23–93. DOI: 10.2903/j. efsa.2011.1930.
- 15. Arvanitoyannis, I. S. (2010). Irradiation of food commodities: techniques, applications, detection,

legislation, safety and consumer opinion. London, UK: Academic Press.

- Piniero, M., & Diaz, L. B. (2007). Improving the safety and quality of fresh fruit and vegetables (FFV): A practical approach. *Acta Hortic.*, 741, 19–24. DOI: 10.17660/ActaHortic.2007.741.1.
- Kume, T., Furuta, M., Todoriki, S., Uenoyama, N., & Kobayashi, Y. (2009). Status of food irradiation in the world. *Radiat. Phys. Chem.*, 78(3), 222–226. DOI: 10.1016/j.radphyschem.2008.09.009.
- Kume, T., & Todoriki, S. (2013). Food irradiation in Asia, the European Union, and the United States: A status update. *Radioisotopes*, 62, 291–299. DOI: 10.3769/radioisotopes.62.291.
- Roberts, P. B. (2014). Food irradiation is safe: Half a century of studies. *Radiat. Phys. Chem.*, 105, 78–82. DOI: 10.1016/j.radphyschem.2014.05.016.
- European Food Safety Authority. (2011). Scientific opinion on the chemical safety of irradiation of food. *EFSA Journal*, 9(10), 23–93. DOI: 10.2903/j. efsa.2011.1930.
- Ihsanullah, I., & Rashid, A. (2017). Current activities in food irradiation as a sanitary and phytosanitary treatment in the Asia and Pacific Region and a comparison with advanced countries. *Food Control*, 72, 345–359. DOI: 10.1016/j.foodcont.2016.03.011.
- 22. European Committee for Standardization. (2001). Foodstuffs – Thermoluminescence detection of irradiated food from which silicate minerals can be isolated. EN 1788:2001. EU, Brussels, Belgium.
- Sanderson, D. C. W., Schreiber, G. A., & Carmichael, L. A. (1991). A European trial of TL detection of irradiated herbs and spices. (Scottish Universities Research Reactor Center Report to BCR).
- 24. Schreiber, G. A., Wagner, U., Leffke, A., Helle, N., Ammon, J., Buchholtz, H. -V., Delincée, H., Estendorfer, S., Fuchs, K., von Grabowski, H. -U., Kruspe, W., Mainczyk, K., Münz, H., Nootenboom, H., Schleich, C., Vreden, N., Wiezorek, C. & Bögl, K. W. (1993). *Thermoluminescence analysis to detect irradiated spices, herbs and spice- and herbs mixtures – an intercomparison study*. Instituts für Sozialmedizin und Epidemiologie des Bundesgesundheitsamtes. Berlin: German Federal Health Office (Bundesgesundheitsamt). (SozEp-Heft 2/1993).
- Schreiber, G. A., Helle, N., & Bögl, K. W. (1995). An inter-laboratory trial on the identification of irradiated spices, herbs and spice-herbs mixtures by thermoluminescence analysis. *J. AOAC Int.*, 78(1), 88–93. DOI: 10.1093/jaoac/78.1.88.
- Sanderson, D. C. W., Slater, C., & Cairns, K. J. (1989). Detection of irradiated food. *Nature*, *340*, 23–24. DOI: 10.1038/340023b0.
- Sanderson, D. C. W., Slater, C., & Cairns, K. J. (1989). Thermoluminescence of foods: Origins and implications for detecting irradiation. *Radiat. Phys. Chem.*, 34(6), 915–924. DOI: 10.1016/1359-0197(89)90329-9.
- Sanderson, D. C. W. (1990). Luminescence detection of irradiated foods. In D. E. Johnston & M. H. Stevenson (Eds.), *Food irradiation and the chemist* (pp. 25–56). Cambridge: The Royal Society of Chemistry.
- 29. Autio, T., & Pinnioja, S. (1990). Identification of irradiated foods by the thermoluminescence of mineral

contamination. Z. Lebensm. Unters. Forsch., 191(3), 177–180. DOI: 10.1007/BF01197616.

- Göksu, H. Y., Regulla, D. F., Hietel, B., & Popp, G. (1990). Thermoluminescent dust for identification of irradiated spices. *Radiat. Prot. Dosim.*, 34(1/4), 319–322. DOI: 10.1093/oxfordjournals.rpd.a080912.
- Delincée, H. (1992). Detection methods for irradiated food. In Proceedings Symposium "Irradiation for the food sector", 13 May 1992 (pp. 24–60). Saint-Hyacinthe, Quebec, Canada: Agriculture Canada.
- Autio, T., & Pinnioja, S. (1993). Identification of irradiated foods by thermoluminescence of contaminating minerals. In M. Leonardi, J. J. Raffi & J. Belliardo (Eds.), *Recent advances on detection of irradiated food* (pp. 183–191). Luxembourg: Commission of the European Communities. (EUR-14315).
- Pinnioja, S., Autio, T., Niemi, E., & Pensala, O. (1993). Import control of irradiated foods by the thermoluminescence method. Z. Lebensm. Unters. Forsch., 196(2), 111–115. DOI: 10.1007/bf01185568.
- Pinnioja, S. (1993). Suitability of the thermoluminescence method for detection of irradiated foods. *Radiat. Phys. Chem.*, 42(1/3), 397–400. DOI: 10.10.1016/0969-806X(93)90274-X.
- 35. Schreiber, G. A., Ziegelmann, B., Quitzsch, G., Helle, N., & Bögl, K. W. (1993). Luminescence techniques to identify the treatment of foods by ionizing radiation. *Food Structure*, 12(4), 385–396. DOI: 1046-705X/93\$5.00+0.00.
- Lozano, I. B., Roman-Lopez, J., Tenopala, J. E., Piña-González, H., Guzman-Castañeda, J. I., & Diaz-Gongora, J. A. I. (2023). Thermoluminescence properties and identification of irradiated cocoa beans during long-term storage. *Appl. Radiat. Isot.*, 191, 110532. DOI: 10.1016/j.apradiso.2022.110532.
- Calderón, T., Rendell, H. M., Beneitez, P., Townsend, P. D., Millan, A., & Wood, R. (1994). Thermoluminescence spectra of inorganic dust from irradiated herbs and spices. *J. Food Sci.*, 59(5), 1070–1071. DOI: 10.1111/j.1365-2621.1994.tb08192.x.
- Ahn, J. -J., Akram, K., Lee, J., Kim, K. -S., & Kwon, J. -H. (2012). Identification of a gamma-irradiated ingredient (garlic powder) in Korean barbeque bauce by thermoluminescence analysis. *J. Food Sci.*, 77(4), C476–C480. DOI: 10.1111/j.1750-3841.2011.02614.x.
- 39. Kim, B. -K., Akram, K., Kim, C. -T., Kang, N. -R., Lee, J. -W., Ryang, J. -H., & Kwon, J. -H. (2012). Identification of low amount of irradiated spices (red pepper, garlic, ginger powder) with luminescence analysis. *Radiat. Phys. Chem.*, 81(8), 1220–1223. DOI: 10.1016/j.radphyschem.2012.01.023.
- Kim, B. -K., Kim, C. -T., Park, S. H., Lee, J. -E., Jeong, H. -S., Kim, C. -Y., Lee, J. -K., Yu, M. -A., & Kwon, J. -H. (2015). Application of thermo-luminescence (TL) method for the identification of food mixtures containing irradiated ingredients. *Food Anal. Methods*, *8*, 718–727. DOI: 10.1007/s12161-014-9928-1.
- Karampiperi, M., Theologitis, S., & Kazakis, N. A. (2022). Thermoluminescence characterization of minerals extracted from dried oregano for retrospective and/or sterilization dosimetry. *Radiat. Meas.*, 158, 106850. DOI: 10.1016/j.radmeas.2022.106850.
- 42. Khan, H. M., & Delincée, H. (1995). Detection of radiation treatment of spices and herbs of Asian

origin using thermoluminescence of mineral contaminants. *Appl. Radiat. Isot.*, 46(10), 1071–1075. DOI: 10.1016/0969-8043(95)00193-H.

- Park, E. -J., Jang, H. -N., Jo, D., Kim, G. -R., & Kwon, J. -H. (2013). Physicochemical quality and luminescence characteristics of gamma-irradiated dried fish products. *Korean J. Food Sci. Technol.*, 45(2), 167–173. DOI: 10.9721/KJFST.2013.45.2.167.
- Arvanitoyannis, I. S., Stratakos, A. C. H., & Mente, E. (2008). Impact of irradiation on fish and seafood shelf life: A comprehensive review of applications and irradiation detection. *Crit. Rev. Food Sci. Nutr.*, 49(1), 68–112. DOI: 10.1080/10408390701764278.
- Sanderson, D. C. W., Carmichael, L. A., Spencer, J. Q., & Naylor, J. D. (1996). Luminescence detection of shellfish. In C. H. McMurray, E. M. Stewart, R. Gray & J. Pearce (Eds.), *Detection methods for irradiated foods current status* (pp. 139–148). Cambridge, UK: Royal Society of Chemistry.
- Pinnioja, S., & Pajo, L. (1995). Thermoluminescence of minerals useful for identification of irradiated seafood. *Radiat. Phys. Chem.*, 46(4/6), 753–756. DOI: 10.1016/0969-806X(95)00255-V.
- Wong, Y. C., Sin, D. W. M., & Yao, W. Y. (2016). Food irradiation and its detection. In L. M. Nollet & F. Toldra (Eds.), *Safety analysis of foods of animal origin* (pp. 663–686). Boca Raton, Florida, US: CRC Press.
- Raffi, J., Fakirian, A., & Lesgards, G. (1994). Comparison between electron spin resonance and thermoluminescence in view of identification of irradiated aromatic herbs. *Ann. Fals. Exp. Chim.*, 87, 125–134.
- Polónia, I., Esteves, M. P., Andrade, M. E., & Empis, J. (1995). Identification of irradiated peppers by electron spin resonance, thermoluminescence and viscosity. *Radiat. Phys. Chem.*, 46(4/6), 757–760. DOI: 10.1016/0969-806X(95)00256-W.
- Sanderson, D. C. W., Carmichael, L. A., & Fisk, S. (2003). Thermoluminescence detection of irradiated fruits and vegetables: International interlaboratory trial. *J. AOAC Int.*, 86(5), 971–975. DOI: 10.1093/ jaoac/86.5.971.
- Schreiber, G. A., Wagner, U., Helle, N., Ammon, J., Buchholtz, H. -V., Delincée, H., Estendorfer, S., von Grabowski, H. -U., Kruspe, W., Mainczyk, K., Münz, H., Schleich, C., Vreden, N., Wiezorek, C., & Bögl, K. W. (1993). *Thermoluminescence analysis to detect irradiated fruit and vegetables – an intercomparison study*. Bericht des Instituts für Sozialmedizin und Epidemiologie des Bundesgesundheitsamtes. Berlin: German Federal Health Office (Bundesgesundheitsamt). (SozEp-Heft 3/1993).
- 52. Marchioni, E., Anklam, E., Chabane, S., Delincée, H., Douifi, L., Hungerbühler, H., Pelleau, Y., Pinnioja, S., Raffi, J., Sanderson, D., & Wagner, U. (1999). Detection by thermoluminescence of an irradiation treatment of five species of dehydrated fruit and vegetables. Report on a CTCPA/AIFLD International Interlaboratory Study. Karlsruhe: Bundesforschungsanstalt für Ernährung. (BFE-R-99-02).
- 53. Akram, K., Ahn, J. -J., Kim, G. -R., & Kwon, J. -H. (2012). Applicability of different analytical methods for the identification of γ-irradiated fresh mushrooms during storage. *Food Sci. Biotechnol.*, 21(2), 573–579. DOI: 10.1007/s10068-012-0073-6.

- Arvanitoyannis, I. S., Stratakos, A. C. H., & Tsarouhas, P. (2009). Irradiation application in vegetables and fruits: A review. *Crit. Rev. Food Sci. Nutr.*, 49(5), 427–462. DOI: 10.1080/10408390802067936.
- Todoriki, S., Kameya, H., Saito, K., & Hagiwara, S. (2014). Detection of commercially irradiated potatoes by thermoluminescence and photostimulated luminescence analyses. *Food Sci. Technol. Res.*, 23(3), 555–561. DOI: 10.3136/fstr.20.555.
- Ahn, J. J., Kim, G. R., Akram, K., Kim, K. S., & Kwon, J. H. (2012). Effect of storage conditions on photostimulated luminescence of irradiated garlic and potatoes. *Food Res. Int.*, 47(2), 315–320. DOI: 10.1016/j.foodres.2011.07.031.
- Ahn, J. J., Kim, G. R., Akram, K., Kim, K. S., & Kwon, J. H. (2012). Luminescence characteristics of minerals separated from irradiated onions during storage under different light conditions. *Radiat. Phys. Chem.*, 81(8), 1215–1219. DOI: 10.1016/j.radphyschem.2012.02.002.
- Yazici, A. N., Bedir, M., Bozkurt, H., & Bozkurt, H. (2008). Thermoluminescence properties of irradiated chickpea and corn. *Nucl. Instrum. Methods Phys. Res. Sect. B-Beam Interact. Mater. Atoms*, 266(4), 613–620. DOI: 10.1016/j.nimb.2007.11.044.
- Khan, H. M., Bhatti, I. A., & Delincée, H. (2002). Thermoluminescence of contaminating minerals for the detection of radiation treatment of dried fruits. *Radiat. Phys. Chem.*, 63(3/6), 403–406. DOI: 10.1016/S0969-806X(01)00630-2.
- Sanderson, D. C. W., Carmichael, L. A., & Naylor, J. D. (1996). Recent advances in thermoluminescence and photostimulated luminescence detection methods for irradiated foods. In C. H. McMurray, E. M. Stewart, R. Gray & J. Pearce (Eds.), *Detection methods for irradiated foods current status* (pp. 124–138). Cambridge, UK: Royal Society of Chemistry.
- Heide, L., Guggenberger, R., & Bögl, K. W. (1990). Application of thermoluminescence measurements to detect irradiated strawberries. *J. Agric. Food Chem.*, 38(12), 2160–2163. DOI: 10.1021/jf00102a012.
- Sillano, O., Román, A., Oeza, A., Rubio, T., & Espinoza, J. (1994). Application of thermoluminescence measurements to detect low dose gamma-irradiated table grapes. *Radiat. Phys. Chem.*, 43(6), 585–588. DOI: 10.1016/0969-806X(94)90172-4.
- Khan, H. M., & Delincée, H. (1995). Detection of irradiation treatment of dates using thermoluminescence of mineral contaminants. *Radiat. Phys. Chem.*, 46(4/6), 717–720. DOI: 10.1016/0969-806X(95)00248-V.
- Khan, H. M., Bhatti, I. A., & Delincée, H. (1998). Identification of irradiated pulses by thermoluminescence of the contaminating minerals. *Radiat. Phys. Chem.*, 52(1/6), 145–149. DOI: 10.1016/S0969-806X(98)00064-4.
- Leffke, A., Helle, N., Linke, B., Bögl, K. W., & Schreiber, G. A. (1993). Studies on detection of irradiated citrus fruit and grains: Germination and some other techniques. In M. Leonardi, J. J. Raffi & J. -J. Belliardo (Eds.), *Recent advances on detection of irradiated food. Proceedings* (pp. 111–121). Luxembourg: Commission of the European Communities. (EUR/14315/en).
- 66. Jo, D., Kim, B. -K., Kausar, T., & Kwon, J. -H. (2008). Study of photostimulated- and thermo-luminescence

characteristics for detecting irradiated kiwifruit. *J. Agric. Food Chem.*, 56(4), 1180–1183. DOI: 10.1021/jf072568y.

- European Committee for Standardization. (2022). Foodstuffs – Detection of irradiated foodstuff containing crystalline sugar by ESR spectroscopy. PN-EN 13708:2022. European Union, Brussels, Belgium.
- Guzik, G. P., Stachowicz, W., & Michalik, J. (2008). Study on stable radicals produced by ionizing radiation in dried fruits and related sugars by electron paramagnetic resonance spectrometry and photostimulated luminescence method – I. D-fructose. *Nukleonika*, 53(Suppl. 2), S89–S94.
- Da Costa, Z. M., Pontuschka, W. M., & Campos, L. L. (2005). A comparative study based on dosimetric properties of different sugars. *Appl. Radiat. Isot.*, 62(2), 331–336. DOI: 10.1016/j.apradiso.2004.08.028.
- Guzik, G. P., Stachowicz, W., & Michalik, J. (2015). Identification of irradiated dried fruits using EPR spectroscopy. *Nukleonika*, 60(3), 627–631. DOI: 10.1515/nuka-2015-0093.
- Guzik, G. P., & Stachowicz, W. (2016). Study on radiation-induced radicals giving rise to stable EPR signal suitable for the detection of irradiation in L-sorbose-containing fruits. *Nukleonika*, 61(4), 461–465. DOI: 10.1515/nuka-2016-0075.
- Guzik, G. P., Stachowicz, W., & Michalik, J. (2019). Study on irradiated D-mannose isolated from cranberry. *Nukleonika*, 64(4), 139–143. DOI: 10.2478/ nuka-2019-0018.
- Yordanov, N. D., Aleksieva, K., & Mansour, I. (2005). Improvement of the EPR detection of irradiated dry plants using microwave saturation and thermal treatment. *Radiat. Phys. Chem.*, 73(1), 55–60. DOI: 10.1016/j.radphyschem.2004.06.008.
- 74. Barea Sanchez, M. (2015). Final report of the intercomparison exercise for quality assurance on TL, PSL and EPR irradiated food detection methods (6th round). Spain: Servicio de Toxicologia Alimentaria et Centro Nacional de Alimentacion.
- Raffi, J., & Angel, J. P. (1989). Electron spin resonance identification of irradiated fruits. *Radiat*. *Phys. Chem.*, 34(6), 891–894. DOI: 10.1016/1359-0197(89)90325-1.
- Raffi, J., Angel, J. P., & Ahmend, S. H. (1991). Electron spin resonance identification of irradiated dates. *Food Technol.*, 3/4, 26–30.
- Karakirova, Y., Yordanov, N. D., De Cooman, H., Vrielinck, H., & Callens, F. (2010). Dosimetric characteristics of different types of saccharides: An EPR and UV spectrometric study. *Radiat. Phys. Chem.*, 79(5), 654–659. DOI: 10.1016/j.radphyschem.2009.12.003.
- Raffi, J., Stachowicz, W., Migdał, W., Barabassy, S., Kalman, B., Yordanov, N., Andrade, E., Prost, M., & Callens, F. (1998). Establishment of an eastern network of laboratories for identification of irradiated foodstuffs. Final Report of Copernicus Concerted Action. CCE. (CIPA-CT94-0134).
- Vanhaelewyn, G., Jansen, B., Pauwels, E., Sagstuen, E., Waroquier, M., & Callens, F. (2004). Experimental and theoretical electron magnetic resonance study on radiation-induced radicals in α-L-sorbose single crystals. *J. Phys. Chem. A*, 108(16), 3308–3314. DOI: 10.1021/jp0378860.

- Vanhaelewyn, G., Lahorte, P., Proft, F., Mondelaers, W., Geerlings, P., & Callens, F. (2001). Electron magnetic resonance study of stable radicals in irradiated D-fructose single crystals. *J. Phys. Chem. Chem. Phys.*, *3*, 1729–1735. DOI: 10.1039/B008248L.
- International Organization for Standardization. (2013). Practice for use of the alanine-EPR dosimetry system. ISO/ASTM 51607:2013. Geneva, Switzerland.